

1 **1. The dark adapted red flash ERG extended protocol.**

2

3 **2. Scope and applications**

4

5 The ISCEV ERG Standard [1] describes a minimum protocol to test rod and cone system function in
6 the outer and inner retina. This extended protocol proposes the inclusion of red flashes under dark
7 adapted (DA) conditions. The DA red flash ERG can be used to distinguish the function of DA rod and
8 cone systems and can help determine the origins of abnormalities seen in the Standard flash ERGs
9 which may be important for accurate characterization of retinal function and to establish some
10 diagnoses.

11 It is well established that the normal cone system contributes to the full-field ERG under DA as well
12 as light adapted (LA) conditions. This occurs in DA ERGs evoked by flash strengths greater than 0.1
13 cd.s.m^{-2} , [2] including the ISCEV standard DA 3 (“combined rod cone”) and DA 10 (“strong flash”)
14 ERGs. Early investigations revealed the contribution of DA cones in the ERG waveform by using
15 colored flashes that exploited differences in the spectral sensitivities of rods and cones [3-5]. These
16 studies showed that the DA ERG waveform to a red flash has two positive peaks. The first, named
17 the x-wave, occurred within 30-50ms and was attributed to DA cone activity. The x-wave was
18 followed by a rod-mediated b-wave [3]. The x-wave is larger than the b-wave during the early stages
19 of dark adaptation when the rod system threshold is high. As dark adaptation proceeds the x- and b-
20 wave amplitudes become similar and finally the b-wave exceeds the x-wave [6].

21 The DA red flash ERG has several clinical applications and circumstances and diagnoses that may
22 benefit from testing are outlined below:

23 a) The DA red flashes are usually well tolerated by patients of all ages, and the test is therefore
24 useful if photophobia or photo-aversion confounds the recording of standard LA ERGs. This can
25 occur in the presence of cone dysfunction, but also for example in the presence of media opacity or
26 strong Bell’s phenomenon.

27 c) In cases of generalized cone system dysfunction such as rod- and S-cone monochromacy and cone
28 dystrophy, the DA red flash ERG x-wave may be undetectable, markedly attenuated and/or
29 delayed[7-9].

30 c) In cases of generalized retinal dysfunction the relative involvement of the DA red flash ERG x-wave
31 and b-wave may suggest predominant dysfunction of cone or rod systems, not always obvious by
32 comparing standard DA and LA ERGs.

33 d) In cases of severe or selective rod dysfunction the DA red flash ERG can help determine the causes
34 of abnormal or residual DA bright flash ERGs. This occurs for example in vitamin A deficiency [13],
35 fundus albipunctatus (RDH5-retinopathy) [10, 11] and Oguchi disease (SAG- or GRK1- retinopathy)
36 [6] and in some cases of rod-cone dystrophy including early stages of Bothnia dystrophy (RLBP1-
37 retinopathy). In these disorders the DA 3 and DA 10 ERGs have reduced a-waves indicating rod
38 photoreceptor dysfunction, but there may also be reduction in the b:a ratio. The reduced b:a ratio
39 may arise from strong stimulation of the relatively preserved DA cone system, analogous to the
40 photopic hill phenomenon, and produces a b-wave which resembles the waveform of the x-wave.

41 e) “Bradyopsia” (RGS9- and R9AP-retinopathy). The DA red flash ERG is normal, but LA cone-
42 mediated ERGs are extinguished by repetitive flashes [9, 10]. The combination of a preserved DA red
43 flash ERG x-wave and undetectable or severely abnormal standard LA ERGs is pathognomonic for the
44 disorder.

45 e) The red flash ERG has been used to detect color vision deficiencies and has been reported to be
46 absent [8, 12] or subnormal [9] in protanopia. The implication is that around 1/100 males would
47 have an absent red flash ERG although this has not been established for an ISCEV DA red flash ERG
48 extended protocol.

49

50 **3. Identification**

51 Red Flash ERG v5 061117 Corresponding author: Dorothy A Thompson.

52 Co-authors: Kaoru Fujinami, Ruth Hamilton, Ido Perlman, Anthony G. Robson

53

54 **4. Patient population**

55 Patients of all ages, referred for investigation of possible retinal dysfunction, retinal dystrophy,
56 generalized cone or rod system dysfunction or patients with photophobia may benefit from the DA
57 red flash ERG, embedded within the ISCEV standard full-field ERG protocol.

58

59 **5. Technical issues**

60 The DA red flash ERG will follow the specifications of the current ISCEV standard full-field ERG and
61 for most applications may be embedded within the standard protocol [1]. Additional considerations
62 include the following:

63 a) The spectral characteristics of the red flash. Both peak wavelength and bandwidth may affect the
64 DA red flash ERG. Physical filters e.g. Kodak Wratten filters 26 (dominant wavelength 619nm) or 29
65 (dominant wavelength 630nm) were used in many older studies, but have been largely superseded
66 by LEDs e.g. peak wavelengths 635nm or 655nm It is noted that peak wavelengths shorter than
67 620nm may be perceived as orange and that for wavelengths longer than 650nm, waveforms have
68 been reported with a third positive wave, later than the rod b-wave [6].

69 b) The units of flash strength. The relative (effective) strength of a colored flash depends upon the
70 adaptation and hence spectral sensitivity of the eye. Absolute measures are radiant energy, but for
71 uniformity of clinical use and consistency with other flash stimuli, it is suggested to use photometric
72 units defined in phot cd.s.m⁻².

73 c) Duration of dark adaptation. The choice of dark adaptation duration and flash strength depends
74 upon one of three aims (Figure 1):

75 i) To isolate the cone-mediated x-wave (peak time 30–50ms): short dark-adaptation of around 5
76 minutes reveals the x-wave before it is masked by full development of the later rod-mediated b-
77 wave [6, 7, 13].

78 ii) To separate the x- and b-wave peak times: if an ISCEV Standard period of at least 20 minutes dark
79 adaptation is used, weaker red flash strengths of around 0.03–0.3 cd.s.m⁻² allow maximum
80 separation in time of the cone- and rod-mediated components.

81 iii) To match the amplitudes of the DA red flash ERG b-wave with the ISCEV Standard DA 0.01 ERG
82 (rod ERG) b-wave: stronger red flashes have been used. This red flash strength may be subject and
83 age specific if defined in this way e.g. [17]. Further subtraction analysis is beyond the scope of this
84 proposal and can be problematic e.g. [18].

85 d) Frequency of red flash presentation. The inter-stimulus interval will influence the light adaption of
86 the retina and shape of the DA red flash ERG waveform [14]. A flash rate of 1 per s does not result in
87 diminishing ERG amplitudes [19], but the effects of faster flash rates are not fully established and
88 require further investigation. The ISCEV standard for the DA 0.01 ERG is greater or equal to 1 flash
89 every 2s and a similar frequency may be appropriate for flash strengths that elicit responses of
90 similar amplitude to the DA 0.01 ERG.

91

92 **6. Calibration**

93 Calibration is in accordance with the ISCEV ERG standard [1]. A spectral photometer is required to
94 determine the spectral characteristics of the red flash. Stimulators may use different combinations
95 of LEDs for different flash strengths, so equal spectral characteristics should not be assumed.

96

97 **7. Protocol specification**

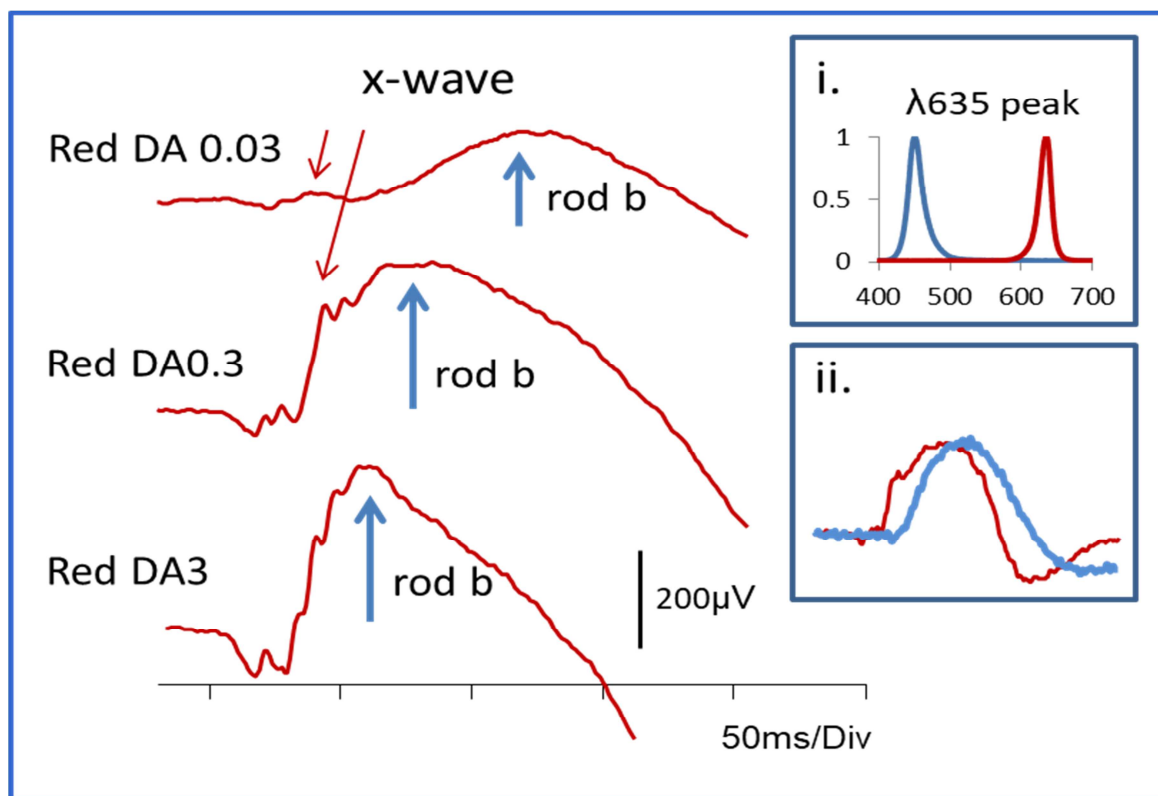
98 Patient preparation follows that for the current ISCEV ERG Standard [1] and the DA red flash ERG
99 may be embedded within the standard ERG protocol. The following additional specifications are
100 suggested.

101 a) Stimulus wavelength. For routine diagnostic applications an LED with a peak wavelength of
102 between 635nm (figure 1) and 650nm is suggested to allow separation of x- and b-waves. If Xenon
103 flashes and filters are used it is suggested to use a dominant wavelength of 619nm (e.g. Wratten 26)
104 or 630nm (e.g. Wratten 29). The peak wavelength and bandwidth of the stimulus and method of
105 generation (optical filter or LED), should be stated.

106 b) Flash strength. It is suggested that a red flash strength of 0.3 cd.s.m⁻² is included. This has been
107 commonly used. This does not preclude the recording of additional red flash ERGs (ranging around
108 0.3 cd.s.m⁻², see figure 2, to account for age, pupillary dilatation etc), but care should be taken to
109 avoid light-adapting the retina and it may be necessary to increase the inter-stimulus interval. If the
110 red flash stimulus is defined according to that required to elicit a DA red flash ERG b-wave of equal
111 or similar amplitude to the DA 0.01 ERG, this should be acknowledged and the corresponding flash
112 stimuli stated in cd.s.m⁻².

113 c) Duration of dark adaptation. A stimulus strength of 0.3 cd.s.m^{-2} may be incorporated within the
114 ISCEV Standard ERG protocol, after a minimum of 20 minutes DA and after the DA 0.01 ERG. There
115 may be specific reasons for recording the DA red flash ERG after shorter periods of DA (see section
116 5c; also to minimize the overall recording time), but care should be taken to avoid significant light
117 adaptation prior to the DA ERGs. Mesopic cone-rod interactions associated with shorter DA may
118 increase the variability of the DA red b-wave amplitude.

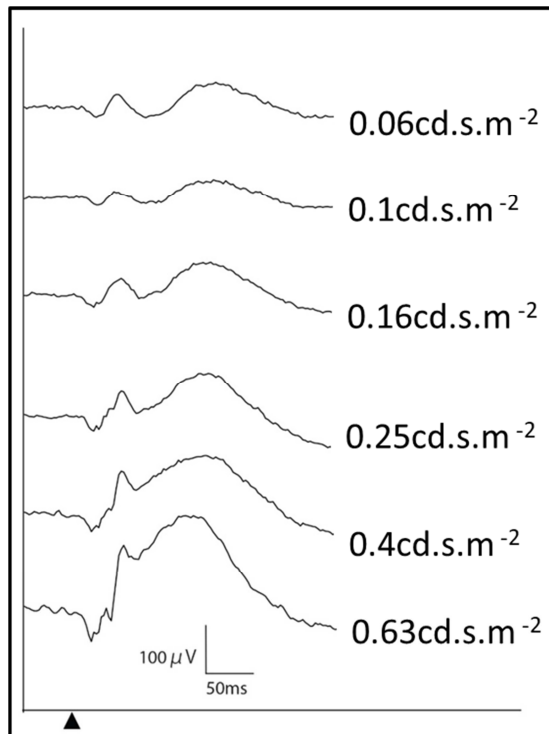
119 d) Frequency of red flash presentation. A flash rate of or between 0.5 and 1 per s is suggested,(i.e.
120 an inter-stimulus interval 1 flash every 2 seconds), but longer inter-stimulus intervals may be needed
121 for stronger red flashes. A maximum rate of 0.5 per s conforms to the current ISCEV Standard for the
122 DA 0.01 ERG.



123

124

125 Figure 1: the change in DA ERG waveform to three red flash strengths is shown after 20 minutes dark
126 adaptation from a Caucasian patient. Note the separation in peak time of the x-wave and b-wave to
127 dim 0.03 cd.s.m^{-2} flashes, the enlargement of the x-wave to 0.3 cd.s.m^{-2} and the merging of x- and b-
128 waves at DA 3 in a control subject. Insert i. shows the spectral characteristics of the red and blue
129 LEDs in the Ganzfeld. Insert ii shows the DA red flash ERG to 0.3 cd.s.m^{-2} in a second subject
130 compared with a DA blue flash ERG of 'scotopically matched' b-wave amplitude, in this case DA blue
131 $0.0003 \text{ cd.s.m}^{-2}$. DA red ERGs shown in red, DA blue flash ERGs shown in blue.



132

133 Figure 2. DA red flash ERGs are shown to a range of flash strengths that includes 0.3cd.s.m^{-2} ,
 134 recorded from a Japanese patient after 20 minute DA $\lambda 650\text{nm}$.

135

136 **8. Response evaluation**

137 Examples of the DA red flash ERG waveforms are shown in figure 1 for different flash strengths
 138 delivered using an LED (peak wavelength 635nm; bandwidth as shown). It is suggested that the DA
 139 red flash ERG parameters are noted as follows: a-wave (if present): earliest trough, amplitude
 140 relative to baseline, peak time relative to flash midpoint: x-wave: peak or shoulder, amplitude
 141 relative to baseline or a-wave trough (if present), peak time relative to flash midpoint: b-wave: peak
 142 following x-wave, similar to DA0.01 (rod) ERG b-wave, amplitude relative to baseline or a-wave
 143 trough (if present), peak time relative to flash midpoint.

144

145 **9. Reporting**

146 Reporting the DA red flash should follow the recommendations of the ISCEV ERG protocol. The flash
 147 stimulus characteristics (LED or filter), peak wavelength or filter specification (e.g. Wratten 26 or 29)
 148 should be stated. The flash strength should be stated. Unless already embedded within the ISCEV
 149 standard ERG protocol, pupil size and duration of dark adaption should be stated. The amplitude of
 150 the a-wave, x-wave and b-wave and their respective time to peaks may be reported along with age-
 151 appropriate laboratory reference data. It is acknowledged that in studies involving ISCEV standard
 152 ERGs it may be sufficiently informative to describe the relative reduction or preservation of x-wave
 153 and/or b-waves relative to each other and normal values.

154

155 **10. References**

156

- 157 1. McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R, et al. ISCEV
158 Standard for full-field clinical electroretinography (2015 update). *Documenta ophthalmologica*
159 *Advances in ophthalmology*. 2015;130(1):1-12. doi: 10.1007/s10633-014-9473-7. PubMed PMID:
160 25502644.
- 161 2. Lim SHO, Y. H. Study of blue and red flash in dark-adapted electroretinogram. *Korean Journal*
162 *of Ophthalmology*. 2005;19(2):106-11. PubMed PMID: 15988925.
- 163 3. Motokawa K MT. Über eine einfachere Untersuchungsmethode und Eigenschaften der
164 Aktionsströme der Netzhaut des Menschen. *Tohoku J Exp Med* 1942;42:114–33. doi:
165 doi:10.1620/tjem.42.114.
- 166 4. Adrian ED. The electric response of the human eye. *J Physiol*. 1945;104(1):84-104. PubMed
167 PMID: 16991672; PubMed Central PMCID: PMCPMC1393520.
- 168 5. Adrian ED. The rod and cone components in the electrical response of the human eye. . *J*
169 *Physiol* 1946;104:84–104. doi: doi:10.1113/jphysiol.1945.sp004109.
- 170 6. Auerbach E, Burian, H.M. . Studies on the photopic-scotopic relationships in the human
171 electroretinogram. *Am J Ophthalmol* 1955;40((5)):42–60.
- 172 7. Kriss A, Jeffrey B, Taylor D. The electroretinogram in infants and young children. *J Clin*
173 *Neurophysiol*. 1992;9(3):373-93. PubMed PMID: 1517405.
- 174 8. Francois JV, G.; De Rouck, A. Pathology of the x-wave of the human electroretinogram. I.
175 Red-blindness and other congenital functional abnormalities. *Br J Ophthalmol*. 1956;40(7):439-43.
176 Epub 1956/07/01. PubMed PMID: 13355951; PubMed Central PMCID: PMCPMC1324669.
- 177 9. Kellner UF, M. H. Color electroretinography. A method for separation of dysfunctions of
178 cones. *Documenta ophthalmologica Advances in ophthalmology*. 1992;80(1):13-23. Epub
179 1992/01/01. PubMed PMID: 1505335.
- 180 10. Michaelides ML, Z.; Rana, N. A.; Richardson, E. C.; Hykin, P. G.; Moore, A. T.; Holder, G. E.;
181 Webster, A. R. Novel mutations and electrophysiologic findings in RGS9- and R9AP-associated retinal
182 dysfunction (Bradyopsia). *Ophthalmology*. 2010;117(1):120-7.e1. PubMed PMID: 19818506.
- 183 11. Robson AGS, P. I.; Sohn, E. H.; Li, Z.; McBain, V. M.; Wright, G.; Moore, A. T.; Webster, A. R.;
184 Holder, G. E., editor Genotype-phenotype variability in patients with clinical and/or
185 electrophysiological features of "fundus albipunctatus". *Documenta Ophthalmologica*; 2011
186 December.
- 187 12. Schubert Von G, Bornschein, H. Beitrag zur Analyse des menschlichen Elektroretinogramms.
188 *Ophthalmologica*. 1952;123((6)):396–412.
- 189 13. Hamilton R, Graham, K. Effect of shorter dark-adaptation on ISCEV standard ERGs and an
190 exploration of the dark-adapted red flash ERG. 53rd Symposium of International Society for Clinical
191 Electrophysiology of Vision (ISCEV), 23–27 June 2015 Ljubljana, Slovenia. Berlin Heidelberg: Springer-
192 Verlag 2015. p. 43.
- 193 14. Verdon WA, Schneck ME, Haegerstrom-Portnoy G. A comparison of three techniques to
194 estimate the human dark-adapted cone electroretinogram. *Vision Res*. 2003;43(19):2089-99.
195 PubMed PMID: 12842161.
- 196 15. Miyake Y. *Electrodiagnosis of retinal disease*. Japan: Springer-Verlag; 2006. 233 p.
- 197 16. Liu X, Liu L, Li H, Xu F, Jiang R, Sui R. RDH5 retinopathy (fundus albipunctatus) with preserved
198 rod function. *Retina*. 2015;35(3):582-9. doi: 10.1097/IAE.0000000000000319. PubMed PMID:
199 25170858.

- 200 17. Mizunoya S, Kuniyoshi K, Arai M, Tahara K, Hirose T. Electroretinogram contact lens
201 electrode with tri-color light-emitting diode. *Acta Ophthalmol Scand*. 2001;79(5):497-500. PubMed
202 PMID: 11594987.
- 203 18. Chen L, Png R, Mathur R, A. C. Scotopic red ERG findings. 53rd Symposium of International
204 Society for Clinical Electrophysiology of Vision (ISCEV), 23–27 June 2015 2015; Ljubljana, Slovenia.
205 Berlin Heidelberg: Springer-Verlag 2015. p. 31.
- 206 19. Lovasik JVK, A. C.; Kergoat, H. Improving the diagnostic power of electroretinography by
207 transient alteration of the ocular perfusion pressure. *Optometry and Vision Science*. 1992;69(2):85-
208 94. PubMed PMID: 1584558.
- 209 20. Chia A PR, Mathur R. , editor Scotopic red response: rod and cone components. International
210 Society for Clinical Electrophysiology of Vision Symposium; 2014; Boston, USA: Doc Ophthalmol
- 211 21. Weleber RG. The effect of age on human cone and rod ganzfeld electroretinograms.
212 *Investigative Ophthalmology & Visual Science*. 1981;20(3):392-9. PubMed PMID: 7203883.
- 213 22. McBain VA EC, Pieris SJ, Supramaniam G, Webster AR, Bird AC, Holder GE. Functional
214 observations in vitamin A deficiency: diagnosis and time course of recovery. *Eye* 2007;21:367–76. .
215 doi: doi:10.1038/sj.eye.6702212.
- 216 23. Sergouniotis PI, Sohn EH, Li Z, McBain VA, Wright GA, Moore AT, et al. Phenotypic variability
217 in RDH5 retinopathy (Fundus Albipunctatus). *Ophthalmology*. 2011;118(8):1661-70. doi:
218 10.1016/j.ophtha.2010.12.031. PubMed PMID: 21529959.
- 219 24. Cheng JYC LC, Yong VHK, Mathur R, Aung T, Vithana EN Bradyopsia in an Asian man. . *Arch*
220 *Ophthalmol* 2007;125:1138–40. doi: doi:10.1001/archopht.125.8.1138.
- 221 25. Vincent A, Robson AG, Holder GE. Pathognomonic (diagnostic) ERGs. A review and update.
222 *Retina*. 2013;33(1):5-12. doi: 10.1097/IAE.0b013e31827e2306. PubMed PMID: 23263253.
- 223 26. Creel DJ, editor Scotopic dim blue and red ERG stimuli. *Documenta Ophthalmologica*; 2013
224 October.
- 225 27. Iijima HY, S. [Adaptational changes in cone electroretinograms in man]. *Nippon Ganka*
226 *Gakkai Zasshi - Acta Societatis Ophthalmologicae Japonicae*. 1990;94(11):987-92. PubMed PMID:
227 2075875.
- 228 28. Lovasik JVK, H. Influence of transiently altered retinal vascular perfusion pressure on
229 rod/cone contributions to scotopic oscillatory potentials. *Ophthalmic & Physiological Optics*.
230 1991;11(4):370-80. PubMed PMID: 1771074.
- 231 29. Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, et al. Preferred reporting
232 items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Systematic*
233 *Reviews*. 2015;4(1):1. doi: 10.1186/2046-4053-4-1. PubMed PMID: PMC4320440.

234

235 **Part B. Justification for the protocol details and description of the consultation process**

236 A systematic review of stimulus parameters and uses of the DA red flash ERG are summarized and
237 tabulated below, with a second table summarizing some of the parameters currently in use by ISCEV
238 members. Feedback was incorporated following presentation of the draft protocol at the BrisCEV
239 eptember 2017 and ISCEV October 2017.

240 Our review highlights two evidence gaps:

- 241 1) Which LED red flash strengths are optimal for different durations of dark adaptation, to
242 visualize the x-wave and maximize the diagnostic utility of x- and b-waves.
- 243 2) What happens to the DA red flash ERG in protanopia produced by the suggested extended
244 protocol.

245

246 **REVIEW OF EVIDENCE**

247 Published specifications are tabulated below (Table1). In summary, red flash strengths range
 248 between 0.05, 0.1 [15], 0.5 [16], 0.17cd.m⁻² [17], 0.25 [18], 2.37 [19], 2.4 [2], 1.5 and 2.5 cd.s/m²
 249 [20] at 20 minutes DA. When Grass strobes have been used to deliver the red flash the range of
 250 intensity settings 1, 4, 8 and 16 have been used, e.g. gr4 white PS22 ~ 3.7 x10⁵ candles [7] or gr4 +
 251 Wratten 26 filter = 0.02 Log μJoule/cm²-steradian [21]. Sometimes no numerical value, nor
 252 wavelength, is stated: some studies reporting clinical use of the red flash ERG describe flash strength
 253 “such that in a normal subject the amplitude of the rod component to the red flash is equivalent to
 254 that of the rod-specific response to a dim white flash (dark-adapted 0.01 cd s m⁻²)”, without
 255 providing a value [22-25]. Others suggest the red flash luminance is empirically set to achieve
 256 ~200μV amplitude scotopic b-wave [26].

257 TABLE 1 Published stimulus details are tabulated, where available indication of normal response are
 258 stated or derived from published figures.

data	Peak λ	Flash strength	DA duration	LED/Xenon
[6] Auerbach & Burian 1955	Wratten 29 635nm Wratten 70 650nm	6 & 12 cd.s.m ⁻²	5 mins	Xenon @30cm
[8] Francois et al 1956	Neon 570 nm	0.1 Joule		Neon 0.2s (orange)
	x-wave 25-60uV@40ms			
[27] Iiyami & Yamaguchi 1990	Wratten 29 blocks below 600nm	86-112cd.s/m ²	30 minutes	
[9] Kellner & Foerster 1992	Wratten 29 623nm	?not stated		Xenon in Ganzfeld
[28] Lovasik et al 1992	Wratten 26 >600 nm	2.37 cd.s.m ⁻²	Not stated	Xenon in Ganzfeld
<i>From figure</i>		90uV@50ms		
[17] Mizunoya et al 2001	LED 660nm	0.17 cd.s.m ⁻²	20 minutes	C/L ganzfeld
[14] Verdon et al 2001	Wratten 26 >600 nm			Xenon Ganzfeld
<i>From figure</i>		@40ms		
[2] Lim and Ohn 2005	Wratten 26 =605nm (Scot match -14Db blue)	2.4 cd.s.m ⁻²	45 minutes	Xenon Ganzfeld
<i>Control data</i>		172.4μV@46ms		N=52 adult
[21] Weleber 1981	Wratten 26 >600 nm	Gr1, 4 and 16	30 mins	Xenon in Ganzfeld
<i>Control data</i>	BA C/L	gr1 = 50μV (25-75) @40-50ms gr4 = 150μV@50ms		N=24 adult

		gr16 = 325µV (200-400) @ 50ms		
[7] Kriss et al 1992	Grass red peak 670nm	Gr4	5 minutes	Grass @30cm
<i>Control data</i>	Skin electrode	14.3 (SD4.9) µV@ 40.4ms (SD2.6) lower limit 4.5µV @46.9		N=30 over 5m and adult
[18] Chen et al 2015 abs	Espion colour dome 635nm	0.25 cd.s/m ²	20 mins	LED
<i>Control data</i>	a-wave x-wave b-wave	17.6µV@19.8ms 64.0µV@50.7ms 68µV@72.9ms		N= 37 adult
[13]Hamilton & Graham 2015 abs	Skin electrode	1.5 cd.s/m ²	20 mins	N=16 adults
<i>Control data</i>	DA 1.5 cd.s/m ² scotopically matched DA 0.01 rod b-wave 1-5 mins better defined X-wave			

259

260 Of interest, the DA red flash ERG to different λ of the filters were investigated in early studies and
 261 using a very deep red, Wratten 70, (> 650nm), produced a third positive wave, later than the rod b-
 262 wave which has been regarded as specific for chromatic red flash [6], seen also in 660nm red flash
 263 figure of [17].

264 TABLE 2. Specifications used by ISCEV members - Personal communications

ISCEV LABS	Peak λ	Flash strength	DA duration	LED/Xenon
GOSH UK	635nm	0.3 and 2.25 cd.s.m ⁻²	20 mins	LED
MEH UK	645nm	0.2 & 0.3 cd.s.m ⁻²	20 mins	LED
JAPAN MIE& NISO	650nm	0.06 & 0.40 cd.s.m ⁻² range 0.06-20 cd.s.m ⁻²	20 mins	LED
	Watts/steroradian/m ² /nm (-1.6, -1.2, -0.8, -0.4, 0, 0.4, 0.8, 1.3 log cds/m ²)			
GOSH scot match to 0.01 b-wave Control data	a-wave: b-wave: 8-13 yrs	16uV@49ms median (5 th -95 th 37-92uV@13-29ms) 305uV@52ms median (5 th -95 th 179-650uV@45-72ms)		

265

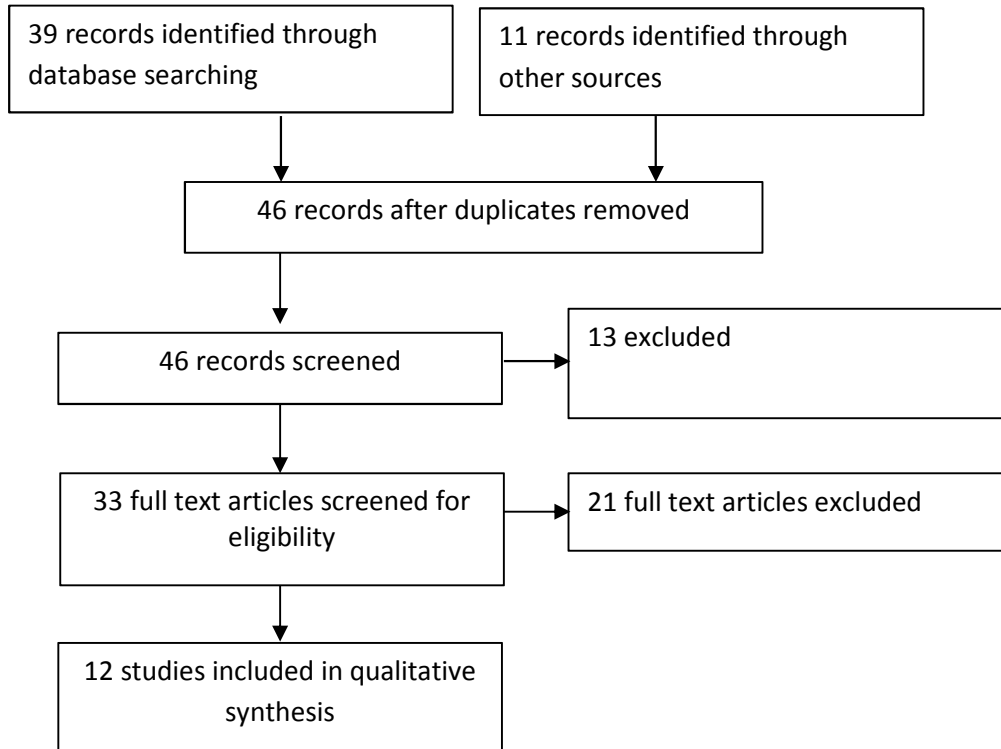
266

267 We followed the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA)
 268 [29] when writing this report. The search strategy aimed to identify reports of scotopic red flash
 269 ERGs in order to extract stimulus parameters of wavelength, flash strength, stimulus duration,
 270 temporal frequency, dark adaptation period and amass evidence of its clinical application and range
 271 of response expected in normal and clinical cases.

272 A systematic literature review was performed to find publications that reported the scotopic red
 273 flash ERG from the period January 1942 to 10/04/2017 using Medline, EMBASE and Cochrane
 274 reviews. The search strategy is shown in appendix 1. Exclusion criteria were animal studies and
 275 absence of any stimulus specification.

276 **Data Collection**

277 The search resulted in 39 items that were exported to Endnote XI. A further 11 items were identified
278 and after duplicates were removed, 46 papers were screened. 30 were eligible for further review
279 and underwent full review, after which 21 were excluded, mostly because they mentioned red flash
280 without any stimulus specification of flash strength or wavelength or because the dark-adapted cone
281 ERG a-wave was mentioned without discussing x- or b-wave.



282

283

284 **APPENDIX 1 Search strategy**

1. exp electroretinography/ (15076)
2. ganzfeld.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
3. ganzfeld stimul*.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
4. full field ERG.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
5. full field stimul*.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
6. exp retina cone/ (4073)
7. cone.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
8. cone photoreceptor.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
9. red flash.mp. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]
10. x-wave.mp. [mp=title, abstract, heading word, drug trade name, original title, device

manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]

11. 1 or 2 or 3 or 4 or 5

12. 6 or 7 or 8

13. 9 or 10

14. 11 and 12 and 13

15. limit 14 to human