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ISCEV Standard for Clinical Electro-oculography (2017 update)

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This update was approved by ISCEV. This document is available on the ISCEV website: <http://www.iscev.org>.

The authors represent the International Society for Clinical Electrophysiology of Vision.

## 23 **Abstract**

24 The clinical electro-oculogram (EOG) is an electrophysiological test of the outer  
25 retina and retinal pigment epithelium (RPE) in which changes in the electrical  
26 potential across the RPE are recorded during successive periods of dark and light  
27 adaptation. This document presents the 2017 EOG Standard from the International  
28 Society for Clinical Electrophysiology of Vision (ISCEV: [www.iscev.org](http://www.iscev.org)). This  
29 standard has been reorganized and updated to include an explanation of the  
30 mechanism of the EOG, but without substantive changes to the testing protocol from  
31 the previous version published in 2011. It describes methods for recording the EOG in  
32 clinical applications and gives detailed guidance on technical requirements, practical  
33 issues, and reporting of results with the main clinical measure (the Arden ratio) now  
34 termed the **Light Peak:Dark Trough ratio (LP:DT<sub>ratio</sub>)**. The standard is intended to  
35 promote consistent quality of testing and reporting within and between clinical  
36 centers.

37

38 **Key words** ISCEV standards, Clinical Electrophysiology, Electro-oculogram (EOG),  
39 Arden ratio, Light Peak:Dark Trough ratio, Light adaptation, Retinal pigment  
40 epithelium (RPE), Fast Oscillation (FO)

41

## 42 **Abbreviations**

43 DT – Dark Trough

44 EOG – Electro-oculogram

45 ER – Endoplasmic Reticulum

46 ERG – Electro-retinogram

47 FO – Fast Oscillation

48 ISCEV – International Society for Clinical Electrophysiology of Vision

49 LP – Light Peak

50 LP:DT<sub>ratio</sub> – Light Peak:Dark Trough ratio

51 RPE – Retinal Pigment Epithelium

52 TEP – Trans Epithelial Potential

53 This Standard is one of a series of ISCEV Standards and Guidelines for  
54 clinical electrophysiology of vision [1-7]. Current standards are listed on the ISCEV  
55 website [8]. This Standard supersedes previous versions of the ISCEV Standard for  
56 Clinical Electro-oculography which was first published in 1993 [1] and subsequently  
57 revised most recently in 2011 [2]. This Standard contains updated information on the  
58 mechanism and reporting of the EOG and has been reorganized for greater clarity and  
59 for consistency with other ISCEV Standards, without substantive changes to the 2011  
60 testing protocol.

61 This ISCEV Standard describes basic procedures that allow reproducible  
62 recordings that are comparable across laboratories. It is intended that the ISCEV  
63 Standard EOG protocol be used widely, but not to the exclusion of other tests or  
64 protocols that are not covered by this Standard. Electrophysiologists are encouraged  
65 to extend the EOG as required when clinically relevant to maximize the diagnostic  
66 value of the EOG and fast oscillation (FO) recordings.

67 Clinical and research users of the EOG are encouraged to use the most recent  
68 Standard, to achieve consistency of results within and between test centers. Reports of  
69 EOG recordings performed in this manner should cite this 2017 Standard. When a  
70 method is used which deviates from the Standard, the deviations should be fully  
71 described.

72

### 73 **The Electro-oculogram**

#### 74 **Electrophysiology of the outer retina in dark and light**

75 There is a difference in electrical potential between the anterior and posterior  
76 of the eye, known as the standing potential of the eye. The standing potential is an  
77 indirect measure of the trans-epithelial potential (TEP) of the retinal pigment  
78 epithelium (RPE). The TEP is equal to the difference in the membrane potential of the  
79 basolateral and the apical membranes, which are electrically isolated through the  
80 tight-junctions of the RPE. Changes in the resistance between the apical and  
81 basolateral membranes or membrane potentials alter the amplitude of the TEP and

82 thus affect the recorded standing potential of the eye. A change in the standing  
83 potential can be induced by retinal adaptation to a change in ambient illumination.

84 There are two clinical tests of the standing potential of the eye. The EOG  
85 utilizes the RPE's response to changing illumination to assess the function of the  
86 outer-retina and RPE. The EOG is recorded during 15 minutes of dark adaptation  
87 followed by 15 minutes of light adaptation. The FO is a different EOG procedure  
88 performed during alternate 1-minute dark and light periods. The changes in the  
89 standing potential for the EOG and the FO are derived from different mechanisms.

90

### 91 **Mechanism of the EOG**

92 During the 15-minute period of dark adaptation there is a fall in the recorded  
93 standing potential typically reaching a minimum at 10–15 minutes, and this is referred  
94 to as the Dark Trough (DT). Following light onset there is an increase in the TEP of  
95 the RPE producing the light-rise of the EOG. The mechanism initiating the light-rise  
96 is unknown but it requires a normally functioning interface between the rod  
97 photoreceptors and RPE. The light-rise is ultimately the result of an increase in  
98 intracellular free calcium, which is released from the endoplasmic reticulum (ER),  
99 regulated by an interaction between ER bestrophin and L-type calcium channels  
100 associated with the basolateral membrane. Intracellular calcium gates the opening of a  
101 basolateral calcium-dependent chloride channel. Increased chloride conductance  
102 depolarizes the basolateral membrane which increases the TEP, recorded as an  
103 increase in the standing potential of the eye. The light-rise normally reaches a  
104 maximum at 7–12 minutes after light onset and is known as the Light Peak (LP). The  
105 LP is the first of several peaks (the damped oscillation) which become progressively  
106 smaller for up to 90 minutes during continued light exposure.

107 The clinical EOG provides an indirect measure of the minimum amplitude of  
108 the standing potential in the dark (at the DT), and then again at its maximum  
109 amplitude in the light (at the LP). This is expressed as the EOG Light Peak to Dark  
110 Trough ratio ( $LP:DT_{ratio}$ ). ISCEV recommends the term  $LP:DT_{ratio}$  when reporting this  
111 result.

112

**113 Mechanism of the FO**

114 The FO is an optional additional test that has a different mechanism to the  
115 clinical EOG owing to the shorter dark and light intervals used. At light onset there is  
116 a fall in potassium in the sub-retinal space that causes a strong outward  
117 hyperpolarizing potassium current across the RPE's apical membrane and is reflected  
118 in the c-wave of the electro-retinogram (ERG). The fall in sub-retinal potassium also  
119 reduces the transport of chloride ions into the RPE. The reduction in chloride ions  
120 causes the basolateral membrane to hyperpolarize and lowers the TEP generating the  
121 trough of the FO 35–45 seconds after light onset. The TEP returns to normal, as ionic  
122 homeostasis is restored and a peak is recorded during the subsequent dark period after  
123 in a further 35–45 seconds. The alternation between dark and light at one minute  
124 intervals establishes a continuous oscillation that is dependent on changes in ionic  
125 permeability at the apical and basal membranes and the electrical coupling between  
126 these membranes by tight junctions.

127

**128 Diseases affecting the light response of the EOG**

129 The LP:DR ratio of the EOG is affected in some diffuse disorders of the RPE  
130 and disorders of the photoreceptor layer of the retina including acquired retinopathies  
131 and retinal dystrophies characterized by rod dysfunction or chorio-retinal atrophy. In  
132 most of these disorders EOG abnormalities are proportional to the severity of rod-  
133 mediated ERG abnormalities and are not of diagnostic importance. Notable  
134 exceptions include disorders of the bestrophin gene (*BEST1*). These include Best  
135 vitelliform macular dystrophy (Best disease), autosomal recessive bestrophinopathy  
136 (ARB), and autosomal dominant vitreoretinopathopathy (ADVIRC). In Best  
137 disease standard, full-field ERGs are usually normal and the LP:DT<sub>ratio</sub> of the EOG  
138 abnormal. An abnormal EOG may distinguish Best disease from other autosomal  
139 dominant retinal disorders with similar fundus features including some cases of adult  
140 vitelliform macular dystrophy and pattern dystrophy. In ARB and ADVIRC the ERG  
141 is often abnormal but the EOG is severely or disproportionately abnormal. An

142 abnormal EOG, not explained by ERG reduction, may also be associated with some  
143 toxic retinopathies.

144

#### 145 **Principles of the clinical EOG measurement**

146 The standing potential of the eye may be assessed using skin electrodes placed  
147 near the outer and inner canthi of each eye to record successive horizontal saccadic  
148 eye movements. The patient tracks alternating lights separated by a fixed angle, to  
149 enable constant eye movement excursions which are recorded as a series of positive  
150 and negative deflections that coincide with ocular rotation. The magnitude of the eye  
151 movement potential (at a fixed angle) is proportional to the standing potential.

152

#### 153 **The Standard Method**

##### 154 **Technologic requirements**

##### 155 **Electrodes**

156 Skin electrodes such as sintered silver–silver chloride, standard silver–silver  
157 chloride or gold cup electrodes are recommended for recording the EOG. The skin  
158 should be prepared by cleaning, and a suitable paste or gel used to ensure good, stable  
159 electrical connection. The electrode-skin contact impedances should be below 5 k $\Omega$  as  
160 measured between 20 and 40 Hz.

161

##### 162 **Stimulator**

163 Full-field (ganzfeld) stimulation must be used to provide uniform luminance over  
164 the entire visual field of the patient. This is usually achieved using a dome or integrating  
165 sphere. It is incumbent on manufacturers and users of to verify that stimulation meets the  
166 full-field requirement of this standard and provides a comfortable head/chin rest for the  
167 patient. Two red fixation lights located 15° left and right of center should be bright enough to  
168 be just visible to the patient during the dark and light phases of the recordings.

169

##### 170 **Light and dark**

171 The adapting dark phase should take place in total darkness, with the fixation  
172 lights dimmed to the minimum necessary to enable fixation for the patient.

173 The adapting light phase requires a white light with a luminance of 100  
174 photopic  $\text{cd.m}^{-2}$  measured at the position of the eye. To account for minor variability  
175 in equipment and calibration the acceptable range within the Standard for the light  
176 adapting background is 90 to 110 photopic  $\text{cd.m}^{-2}$ . Calibration of the ganzfeld  
177 stimulator should be carried out periodically. Modest room lighting may be turned on  
178 during the light phase providing the ambient luminance is less than that in the  
179 ganzfeld.

180 Note that adapting light sources of different types, such as tungsten, halogen,  
181 LED, and fluorescent, have different spectral characteristics and the color may change  
182 with brightness. This makes the definition of standard lighting inherently imprecise,  
183 although for practical purposes most “white” light of the correct luminance will give  
184 similar results.

185

### 186 **Amplification**

187 Amplifiers should have a band pass of either 0 to 30 Hz (DC), or 0.1 to 30 Hz  
188 (AC), to make recordings of saccadic eye movements appear as square waves (Figure  
189 1). For a  $30^\circ$  saccade, the typical EOG amplitude is between 250 and  $1000\mu\text{V}$  with an  
190 essential frequency content of 0 to 30 Hz. To avoid a loss of information, digitizers  
191 should sample the saccades at a rate of 1kHz or higher in each channel. In theory, the  
192 ideal recording technique is DC amplification but this is generally impractical because  
193 of baseline drift. Thus, we recommend AC recording with a 0.1 Hz high pass filter. If  
194 a higher frequency filter is used (e.g. 0.5 Hz), it will distort the square waves, making  
195 identification of overshoot and stepped saccades difficult.

196

197 \_\_\_\_\_ Insert Figure 1 Near Here \_\_\_\_\_

198

199 Manufacturers should allow the examiner to have access to all of the raw data  
200 for each 10 second recording epoch so that the examiner is able to visually inspect the  
201 individual saccadic records to enable accurate cursor placement around any artefacts  
202 to measure the standing potential as shown in Figure 2.

203

204 \_\_\_\_\_ Insert Figure 2 Near Here \_\_\_\_\_

205

206 The operator should be able to visually inspect saccadic recordings as they are  
207 performed to monitor for artefacts including those caused by blinks, inconsistent eye  
208 movements, patient compliance problems, poor electrode contact or amplifier  
209 saturation. Prompt detection allows artefacts to be minimized or eliminated e.g. by  
210 encouragement of the patient, re-application of recording electrodes or by making  
211 necessary adjustments to the amplifier gain.

212

### 213 **Preparation of the patient**

#### 214 **Pupils**

215 The pupils should be dilated before testing and their size or diameter recorded.  
216 If full pupil dilation is impossible or undesirable, an attempt should be made to  
217 increase the adapting luminance so that an equivalent retinal illumination is  
218 approximated. The amount of light passing through the pupil, when measured in  
219 Trolands, is the product of luminance (in  $\text{cd.m}^{-2}$ ) and pupil area (in  $\text{mm}^2$ ). For  
220 example, to produce the same effect upon the retina, twice as much luminance is  
221 required with a 5-mm diameter pupil (roughly  $20 \text{ mm}^2$ ) than with a 7-mm diameter  
222 pupil (roughly  $40 \text{ mm}^2$ ). The report should describe any deviations from the Standard.

223

#### 224 **Electrode placement**

225 After suitable skin preparation, recording electrodes should be placed close to  
226 the canthi of each eye as in Figure 3. The electrodes from each eye are connected to  
227 separate channels of a differential amplifier. A separate electrode should be attached



228 and connected to the ground. Convenient and commonly used ground electrode  
229 positions include the forehead, vertex, mastoid and earlobe. The impedance between  
230 any pair of electrodes should not exceed 5 k $\Omega$ . The electrodes, amplifier and  
231 impedance meter must be approved for medical use.

232

233 \_\_\_\_\_ Insert Figure 3 Near Here \_\_\_\_\_

234

### 235 **Pre-exposure to light**

236 The patient should be in stable indoor lighting for at least 30 minutes before  
237 the test. Indirect ophthalmoscopy, fundus autofluorescence, fundus photography and  
238 other strong illumination changes must be avoided during this period. As near as  
239 practical, the pre-test light exposure should be the same for all patients.

240

### 241 **Instructions to the Patient**

242 The procedure should be explained to the patient, noting that head position  
243 must not change, as this is one common source of artefact and that the eyes should  
244 only move left and right. Patients should be instructed not to anticipate the onset of  
245 the alternation of the fixation lights but to move their eyes only when the lights  
246 change. Practice the procedure with the recording system prior to dark adaptation, to  
247 familiarize the patient with the task and to check on the stability and quality of the  
248 recorded saccades.

249

### 250 **Clinical recording**

#### 251 **Recording of saccades**

252 Fixation lights should alternate once per second, for 10 seconds out of every  
253 minute. The EOG potentials are recorded during each of these 10 second periods.

254 There should be a warning, verbal or automatic, of the impending start of each

255 recording period to ensure readiness of the patient and operator. Auditory cues during  
256 the recordings may be used and can be helpful in patients with restricted visual fields.

257

### 258 **Dark phase**

259 For the dark phase total darkness should be maintained for 15 minutes, except  
260 for the dim fixation lights. EOG recordings should be made once a minute for 10  
261 seconds, as specified above. The operator should have a concurrent view of the  
262 recordings to check for patient compliance, and for errors such as noise or overshoot.

263

### 264 **Light phase**

265 For the light phase a ganzfeld background light of 100 photopic  $\text{cd.m}^{-2}$  should  
266 be turned on to initiate the light phase response and should remain on for the duration  
267 of the light phase. However, the luminance can be increased gradually over a short  
268 period (e.g. 20 seconds) to minimize patient discomfort. Longer ramps (e.g. lasting  
269 minutes), will alter the responses. Continue the recording for 10 seconds out of every  
270 minute (as above) for at least 15 minutes to register the presence or absence of the LP.  
271 If the LP can be clearly identified during the recordings, then the test may be stopped  
272 before 15 minutes. It may be necessary to extend the light phase to fully characterize  
273 abnormal responses but a delay in the LP should be reported. The patient should  
274 remain positioned in the headrest of the stimulator throughout the procedure, with  
275 eyes open to maintain retinal illumination.

276

### 277 **Patient compliance**

278 Patients will have difficulty performing saccadic eye movements if they  
279 cannot fixate reliably because of poor central vision, diplopia or ocular motility  
280 problems (including nystagmus). Patients with diplopia may be advised to look  
281 between the pair of images, or one eye can be patched if the suspected retinal disorder  
282 is binocular. Patients who are very young or those with a physical or learning  
283 disability may not be able to perform the EOG. In young children with suspected Best

284 disease it may be useful to test their parents, since a carrier of Best disease will have  
285 an abnormal light-rise, irrespective of whether the fundus is normal.

286 Patient compliance can vary due to fatigue or inattention. Common problems  
287 include gradual movement of the head back from the stimulator, head turning,  
288 irregular eye movements during the recording or eye closure during the light phase  
289 can be minimized by having a real-time view of the patient's eyes via an infrared  
290 camera. In most cases, gentle coaching and reminders will minimize the effects of  
291 poor compliance.

292

### 293 **Analysis and reporting**

#### 294 **Saccadic amplitude**

295 The EOG amplitudes should be measured in microvolts ( $\mu\text{V}$ ) either manually  
296 or by a computer algorithm after visual inspection. Care must be taken to avoid  
297 measuring the effects of overshoot (see Figure 2) or irregular (artefactual) saccades.  
298 The average of the amplitudes within each 10-second recording epoch should be  
299 measured, excluding outliers, artifacts or responses consistent with poor eye  
300 movement accuracy. If a computer algorithm is used, it is important to ensure that the  
301 values obtained represent true EOG amplitudes and not artefactual records or outliers.  
302 Common causes of unreliability are overshoot of the fixation lights, stepped rather  
303 than smooth saccades, missing saccades, inverse saccades (eyes going in the opposite  
304 way to the fixation lights), and eccentric fixation in which the saccade length switches  
305 between two or more values.

306

#### 307 **Dark trough and light peak: Smoothing**

308 The average EOG amplitude calculated from each 10 second epoch should be  
309 plotted. However, there is always "noise" in physiological recordings, and the goal of  
310 the EOG measurement is to record the underlying DT and LP, rather than the lowest  
311 or highest single values. Thus, the first critical step is that the underlying physiologic  
312 curve be recognized and drawn, in order to derive reliable DT and LP amplitudes.

313 This can be achieved by various methods including visual inspection, by use of a  
314 flexible spline rule, computer-based curve fitting algorithms or weighted averages of  
315 the EOG amplitudes.

316 Figure 4 shows raw data plotted and the subsequent smoothing of the data  
317 points using, in this case, the weighted mean of the recorded EOG amplitudes at each  
318 time point. It is helpful if uncertain or artefactual values can be identified and marked  
319 at the time of recording, so that they can be ignored later when smoothing and curve-  
320 fitting.

321

322 \_\_\_\_\_ Insert Figure 4 Near Here \_\_\_\_\_

323

#### 324 **Light peak to dark trough ratio calculation**

325 The LP:DT<sub>ratio</sub> of the EOG is computed by dividing the smoothed light peak  
326 by the dark trough amplitudes. It is important to note that a normal LP:DT<sub>ratio</sub> does not  
327 imply a normal DT and this amplitude should be reported as a measure of the standing  
328 potential.

329

#### 330 **Reporting**

331 Clinical reports should state the EOG LP:DT<sub>ratio</sub>, the DT amplitude (in  
332 microvolts) and the time from the start of the light phase to the LP (if present). The  
333 type of adapting light source and pupil size should also be reported. The report should  
334 also describe any difficulties encountered during testing that may affect confidence in  
335 the results, such as patient compliance or inconsistent eye movements.

336

#### 337 **Normative data**

338 Each center must establish its own set of normative values for the EOG and  
339 FO. The median value (not the mean) should be used to define reference values and  
340 the actual values on either side of the median that bracket 90 percent of the reference  
341 ranges (5<sup>th</sup> – 95<sup>th</sup> centile) should be determined by direct tabulation of the reported

342 results. The normal LP:DT<sub>ratio</sub> is typically between 1.7 and 4.3 with a LP time ranging  
343 from 7-12 minutes.

344

#### 345 **Interaction between eyes**

346 The EOG potentials from one eye can contaminate the response from the  
347 other. The magnitude of this effect is approximately 15% with electrodes placed on  
348 each side of the nose close to the inner canthi. It rises to about 40% if the electrodes  
349 come close together and touch, becoming equivalent to a common central electrode  
350 on the bridge of the nose. This interaction can give misleading results if an eye is  
351 electrically inactive e.g. in total retinal detachment or absent. This is because the  
352 defective eye will appear to have the same LP:DT<sub>ratio</sub> as the fellow eye, but with a  
353 much smaller standing potential (DT amplitude). If the eyes have similar standing  
354 potentials but a different LP:DT<sub>ratio</sub> then the measured ratio from the better eye is  
355 enhanced at the expense of that from the weaker eye.

356

#### 357 **Deviation from the Standard**

358 This Standard represents a basic and core procedure for the assessment of  
359 generalized function of the RPE/photoreceptor interface. If a center chooses  
360 techniques that vary from the Standard, such as differences in the luminance level for  
361 the adapting light or in the duration of the dark or light adapting intervals, it is critical  
362 to cite this document and specify any deviations. The standing potential of the EOG  
363 can also be used to monitor eye-movements in studies unrelated to retinal and RPE  
364 pathophysiology.

365

#### 366 **Fast Oscillation**

367 The FO has the opposite polarity to the EOG. Light causes a decrease in the  
368 standing potential, while in darkness there is an increase in the standing potential.

369 The FO is recorded using the same technical specifications as the EOG  
370 (amplifier, electrode placement, fixation targets, background luminance and 1 per

371 second saccades). However, the saccades and the recording should be continuous for  
372 the duration of the test. Light and dark are alternated every 60 seconds to induce the  
373 FO, which has a near sinusoidal appearance (Figure 5). The total number of light-dark  
374 intervals should be at least 4 with either 60 or 75-second intervals of light and dark  
375 giving equivalent results. Pre-adaptation does not affect the FO, so this test can be  
376 performed either independently or before the EOG.

377 Figure 5 shows a FO recording formed by dark and light intervals. The FO  
378 amplitude can be calculated from the average of the peak to trough ratios from each of  
379 the dark/light cycles. The time to the peak or trough should be calculated from the  
380 time of light offset or onset respectively and averaged for the number of light/dark  
381 cycles in the recording.

382

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386 approve the final document. We thank Mitchell Brigell, Michael Marmor and Daphne  
387 McCulloch in particular for their constructive input into this document.

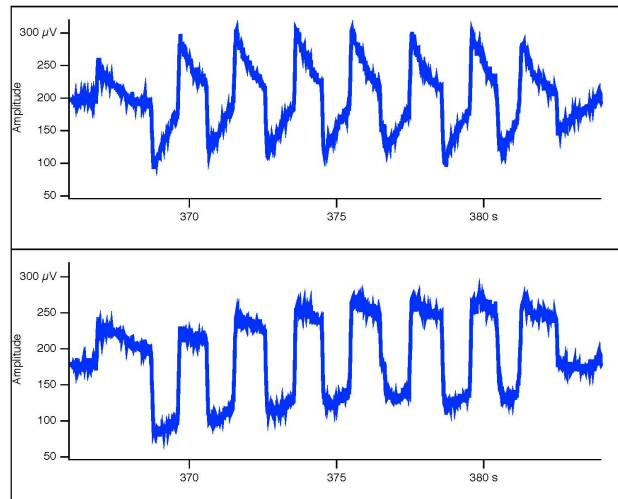
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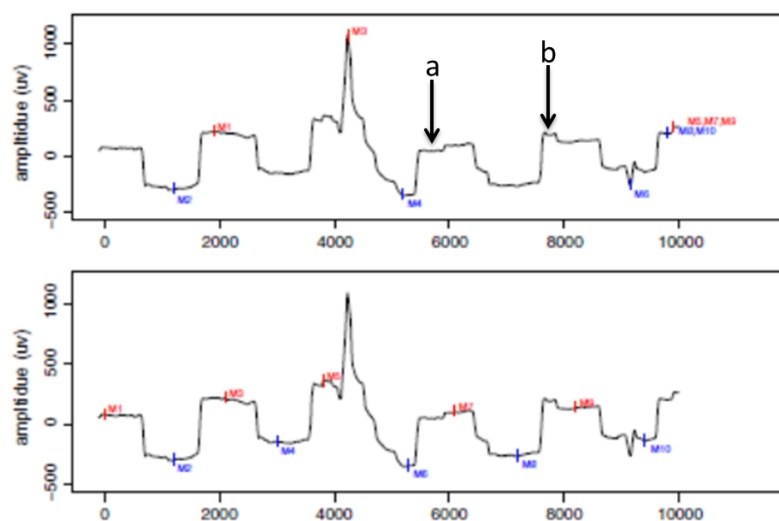
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420

421 **Figures and Captions**

422

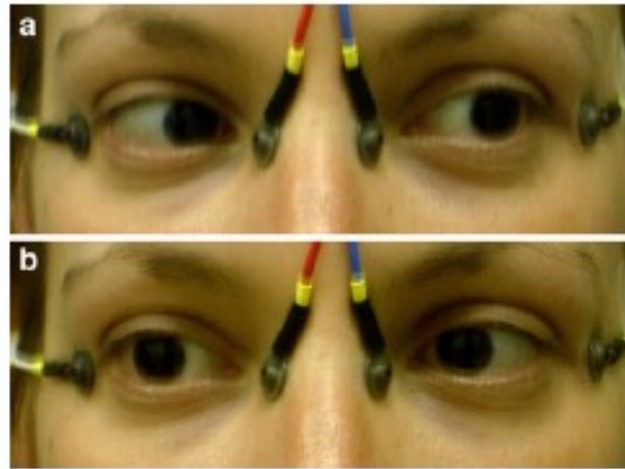
423 **Figure 1** Upper trace shows filtering with 0.1–30 Hz, lower trace with post-hoc DC  
 424 restoration by digital integration, rendering it similar to direct DC recording. DC  
 425 recording (or restoration) makes it easier to perform the plateau measurements.



426

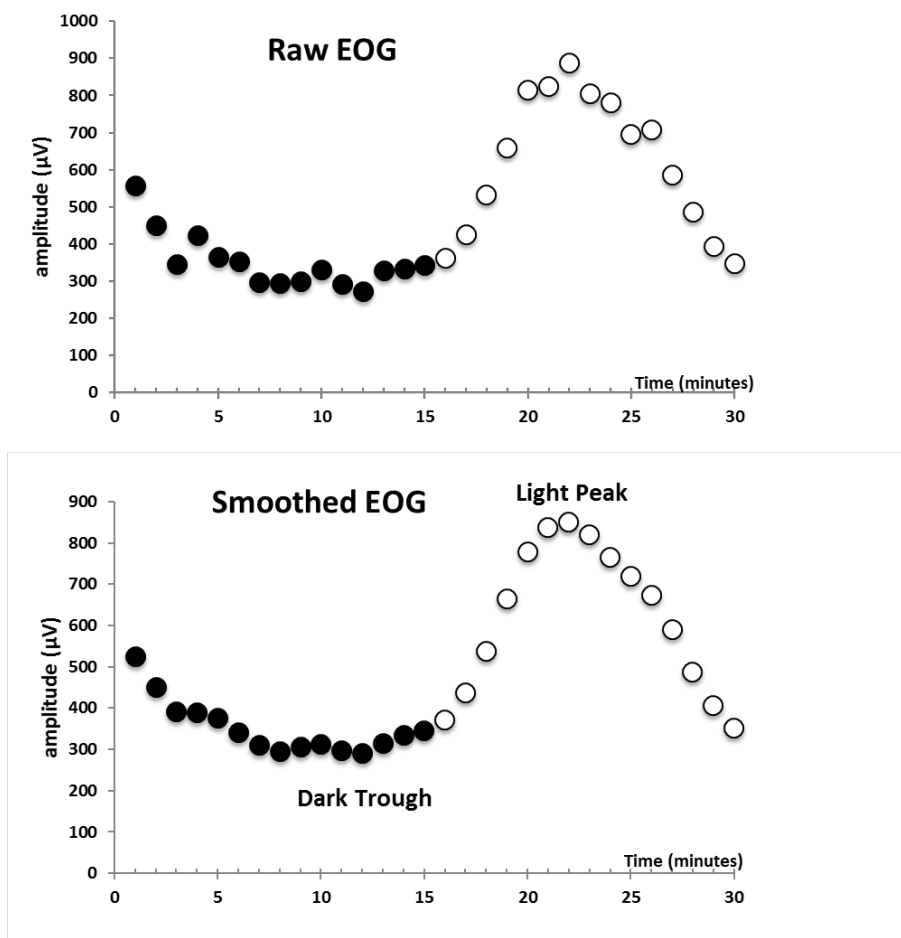
427 **Figure 2** Examples of 10-second saccadic records with a blink artefact at  
 428 approximately 4000 ms. Arrow ‘a’ indicates an initial undershoot and arrow ‘b’ an  
 429 overshoot of the fixation target visible by the step in the plateau of the EOG recording  
 430 (upper trace). The lower trace shows the manual placing of markers at the peak and  
 431 trough of the EOG recording as the eye performs horizontal saccades for ten seconds  
 432 at one second intervals.





433

434 **Figure 3** Recording electrode positions located near the inner and outer canthi of each  
 435 eye. As the eyes perform horizontal saccades: left (a) then right (b) the amplitude of of  
 436 the standing potential is recorded across the active electrodes.

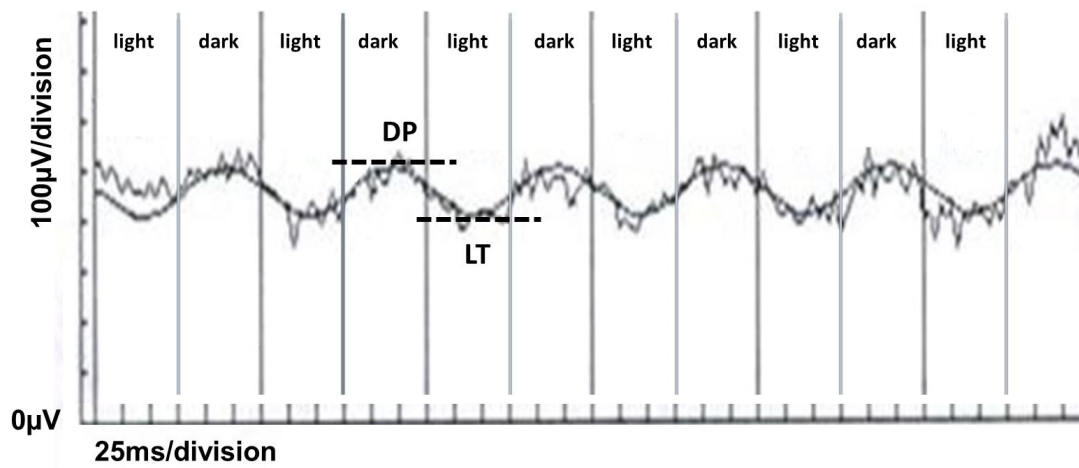


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438

439 **Figure 4** Upper figure shows the raw standing potential values for an EOG with 15  
 440 minutes of dark (black circles) and 15 minutes of light (white circles). Smoothing of  
 441 the data helps to define the DT and LP amplitudes from which to calculate the Light  
 442 Peak:Dark Trough ratio (Lower Figure). Computer algorithms or fitting with a spline  
 443 rule may also be utilized.

444



445

446 **Figure 5.** Typical Fast Oscillation recording from a normal eye with six cycles of  
 447 light and dark intervals (75 seconds each). There is a light trough (LT) and a dark  
 448 peak (DP). In this case amplitude measurements were derived from the fit of the sine  
 449 wave to the raw data. The FO ratio is computed from the amplitude of the DP divided  
 450 by the amplitude of the LT (1.3).